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Phylogeny and taxonomy of the *Ophiostoma piceae* complex and the Dutch elm disease fungi

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Abstract: The *Ophiostoma piceae* complex forms a monophyletic group of insect-dispersed pyrenomycetes with synnemata (*Pesotum*) and micronematous (*Sporothrix*) synanamorphs. Other species of *Ophiostoma* outside of the *O. piceae* complex that form synnema lack the *Sporothrix* state. The nine recognized species within the *O. piceae* complex are delimited by synnema morphology, growth rate at 32 °C, mating reactions and sequences of the internal transcribed spacer (ITS) region of the rDNA operon. Phylogenetic analysis of the ITS region suggests two major clades in the complex, one that causes bluestain in primarily coniferous hosts and the other on primarily hardwood hosts. In the coniferous group are *O. piceae*, *O. carum*, *O. floccosum* and the recently described *O. setosum* (anamorph *Pesotum* *cupulatum* sp. nov.). In the hardwood group are *O. querci*, *O. catonianum*, and the Dutch elm disease fungi: *O. ulmi*, *O. novo-ulmi* and *O. himal-ulmi*. Restriction fragment length polymorphisms of the ITS region are shown to be a convenient diagnostic tool for delimiting these species.

Key Words: Bluestain, Dutch elm disease, *Graphium*, *Leptographium*, *Pesotum*, *Phialographium*, *Sporothrix*

INTRODUCTION

Adaptations for insect dispersal (ascomata and conidiomata with sticky spore drops) have arisen frequently in the evolution of the ascomycetes (Blackwell 1994), and convergence on similar morphologies has greatly confused generic and species delimitations of both teleomorphs and anamorphs (Spatafora and Blackwell 1994). One of the largest groups of insect dispersed pyrenomycetes is the genus *Ophiostoma*, which is now recognized as distinct from *Ceratocystis* based on anamorphs and biology (Hoog and Scheffer 1984, Harrington 1981, 1987). Considerable confusion remains with generic limits for anamorphic fungi dispersed by insects, many of which form droplets of conidia at the apex of compact conidiomata called synnemata (Okada et al 1998). Some synnema-forming species have affinities to *Ophiostoma* H. & P. Sydow, especially to the common sapwood-colonizing species *O. piceae* (Münch) H. & P. Sydow and the Dutch elm disease fungus *O. ulmi* (Buisman) Nannf. (Okada et al 1998).

Synnematous anamorphs of *Ophiostoma* species have been placed in the genus *Graphium* Corda, though it is now thought that *Graphium* species are anamorphs of the Microscales (Okada et al 1998), and the genus name *Pesotum* Crane & Schoknecht is available for the anamorphs of *Ophiostoma* species. *Pesotum* was based on the anamorph of *O. ulmi* and was characterized by the synanemalous anamorph as well as a *Sporothrix* Hektoen & Perkins ex Nicot & Mariat synanamorph (Crane and Schoknecht 1973). *Sporothrix* conidiophores are micronematous, with conidiogenous cells that have prominent denticles at the point of conidium detachment (Hoog 1974, 1993). Secondary conidia are frequently formed from the primary *Sporothrix* conidia, and the name *Hyalodendron* Diddens has been used for such anamorphs. However, the name *Hyalodendron* should be reserved for basidiomycetous anamorphs, and there appears to be little need for distinguishing *Sporothrix* species with secondary conidium production (Hoog 1993). Species of *Hyalorhinocladiella* Upadhyay & Kendrick form micronematous conidiophores similar to those of *Sporothrix* but without prominent denticles (Hoog 1993), and the conidiogenous cells of *Hyalorhinocladiella* resemble those of *Pesotum* species. Synnema forming anamorphs with phialidic conidium production were placed in *Phialographium* Upadhyay & Kendrick, but Hoog and Scheffer (1984) and Wingfield et al (1991) do not distinguish *Phialographium* from *Graphium* *Pesotum* sensu Okada et...
al 1998). *Pesotum*-type conidiophores with a single row of cells for the stipe (mononematous conidiophores) and a series of metulae at the stipe apex are placed in *Leptographium* Lagerb. & Melin (Harrington 1988).

Okada et al. (1998) used *Pesotum* for all synnema forming species with affinities to *Ophiostoma*, even those species with no *Sporothrix* synanamorph. This paper focuses on those *Ophiostoma* species with *Pesotum* anamorphs sensu Crane and Schoknecht (1973), i.e., those that also have *Sporothrix* synanamorphs, such as *O. piceae*. *Ophiostoma piceae* was originally described by Münch (1907) from sapstain in pine and spruce but is now known as a complex of closely related *Ophiostoma* species with pigmented, synnematous anamorphs. Included in the complex are *O. querci* (Brasier and Kirk 1993) and the Dutch elm disease fungi, *O. ulmi*, *O. novo-ulmi* and *O. himal-ulmi* (Brasier and Mehrorda 1995). We found four other species closely tied to the *O. piceae* complex. We used DNA sequences of the internal transcribed spacer (ITS) region of the nuclear rDNA and mating studies to identify lineages and potential intersterility groups, and morphological and physiological comparisons were used to delimit species in the complex.

**MATERIALS AND METHODS**

**Isolates.**—The isolates most intensively studied and considered as part of the *O. piceae* complex are listed in Table 1. Isolates are maintained in the collection at Iowa State University. For morphological comparisons, isolates were grown at room temperature (21–24 C) and lighting for 10–14 d on malt extract agar (MEA, 1.5% Difco malt extract and 2.0% agar) or pine twig medium (MEA with a section of debarked *Pinus strobus* twig added to the molten agar medium, Harrington 1992). Isolates grown for DNA extraction were cultured on MYE broth (2% malt extract, 1% yeast extract).

**Temperature growth assay.**—Isolates were grown at 32 C on MEA because growth at this temperature was shown to distinguish *O. piceae* from *O. querci* (Brasier and Stephens 1993). Plates were inoculated using a plug of agar and mycelium (#1 cork borer, from the advancing margin of a colony) placed upside down in the middle of the plate. The plates were incubated upside down in the dark. After 7 d, the plates were removed from the incubator and two radial measurements taken for each plate. The assay was repeated twice at separate times, and the mean extent of radial growth determined from the three separate experiments.

**Pairings.**—Tests for sexual compatibility were conducted on MEA with pine twigs. Two methods of pairing were used. In most cases, a single isolate was grown on the agar medium for 1 wk and then spermated with a conidial/mycelium slurry (Brasier and Kirk 1993, Harrington and McNew 1997). In other cases, two isolates were co-inoculated with colonized agar plugs placed ca 5 mm apart. Single ascospore strains were obtained from an ascospore mass at the tip of a perithecium by suspending the mass in a light oil and streaking onto MEA plates (Harrington and McNew 1997).

**PCR, DNA sequencing, and RFLP analysis.**—The primers ITS5-F (5’-CTTGGTCAATTAGAGGAGTAA-3’) and ITS4 (5’-TCCTCGATTGATGATGC-3’) (Gardes and Bruns 1993, White et al. 1990) were used to amplify the ITS region. Either extracted DNA (DeScenzo and Harrington 1994) at 10–100 ng per reaction or scraped mycelium with spores (Harrington and Wingfield 1995) were used as template for the polymerase chain reaction (PCR). The reaction mixture (100 μL final volume) contained 2.5 units Taq DNA polymerase (Promega Inc., Madison, Wisconsin), 1X PCR reaction buffer, 1.5 mM MgCl₂, 200 μM dNTPs, 5% (V/V) DMSO, and 0.25 or 0.50 μL of each primer. Reaction mixtures were placed into a preheated (4 C) thermocycler (MJ Research, Inc., Watertown, Massachusetts). Cycling conditions were an initial denaturation at 95 C for 95 s, followed by 35 cycles of 52 C for 40 s (annealing), 72 C for 2 min (extension), and 95 C for 35 s (denaturation). There was a final extension step of 10 min at 72 C.

Amplicons were either sequenced using the primers ITS5-F and ITS4 or were digested with restriction enzymes following the manufacturer’s recommendations. Sequencing was performed on an ABI PRISM 377 Genetic Analyzer (Perkin-Elmer Inc., USA) at the DNA Synthesis and Sequencing Facility (Iowa State University, Ames, Iowa) after purification using QiAquick PCR purification Kits (Qiagen Inc., USA) or Microcon-100 Microcentrators (Amicon, Inc., USA).

The ITS sequences of 58 isolates of taxa in the *O. piceae* complex and an outgroup taxon (isolate C327 of *Ophiostoma ips* from New York, USA) were aligned manually and analyzed using PAUP 4.0 (Swofford 1998). A total of 616 unordered characters, including gaps, were utilized, with gaps considered a “fifth base” and all characters with equal weight. Sixty-four of the characters were parsimony-informative, and 491 characters were constant. Heuristic searches used stepwise (simple) addition and tree-bisection-reconnection. Bootstrap support (Felsenstein 1985) was determined from 1000 replications.

Polymorphisms in restriction sites of the ITS region were identified among species in the *O. piceae* complex using the online tool Webcut 2.0 (http://www.cssi.com/firstmarket/cutter/cut2.html; Max Heiman, copyright 1997). The restriction enzyme *Hae* III (Gibco BRL, Inc., USA) produced the greatest number of polymorphisms among species; however, some species showed similar RFLP patterns with this enzyme. The restriction enzyme *Dde* I (Gibco BRL, Inc., USA) was used to differentiate some taxa. For either enzyme, the unpurified PCR product (17.5 μL) was combined with 2.0 μL of 10X buffer (supplied with the enzyme) and 0.5 μL of restriction enzyme (5 units). Digestion was allowed to proceed for 1–16 h at the appropriate incubation temperature for the enzyme employed. Restriction fragments and a 100-bp ladder (Gibco BRL, Inc., USA) were separated by electrophoresis for 3 h at 75 volts in 2% agarose gels (Bio-Rad, Inc., USA) with a TBE
RESULTS

A wide range of synnema morphologies was seen among the isolates studied. All isolates forming both synnema and Sporothrix states were tolerant of cycloheximide (Harrington 1981) and had similar ITS sequences. The ITS sequence analysis grouped these isolates with bona fide isolates of O. piceae and O. querci (Fig. 1). Nine species in the O. piceae complex are recognized based on the ITS sequences (Fig. 1), mating reactions (Table I), cultural characteristics, and morphology (Figs. 2–36, Table II). Other synnema-forming species, including Ophiostoma species excluded from the O. piceae complex (Table III) and isolates of G. penicillioides, the type species for Graphium, did not form a Sporothrix synanamorph (Figs. 37–48). Their ITS sequences, where available, aligned poorly with sequences of O. piceae and O. querci.

Culture characteristics.—The distinguishing features of the species recognized in the O. piceae complex are listed in Table II. Morphological features are determined on MEA or MEA with pine twig medium, and a relatively weak medium is needed to discern these characteristics. Aroma is useful for distinguishing O. querci and O. setosum from the other species, and this feature is also best determined on MEA. Growth at 32°C is a distinguishing feature for species in the complex (Table I), but accurate temperature control is needed. Protoperithecia, which are distinctly light-brown in O. querci, are most reliably produced 1 wk after a conidial mass from a synnema is streaked over MEA or MEA with pine twig medium. The concentric rings of aerial mycelium that are diagnostically of the Dutch elm disease fungi (Brasier and Mehrtra 1995), O. querci, and O. setosum are also best seen on MEA.

Phylogenetic analyses.—Most of the sequences of the ITS region of other Ophiostoma species [e.g., O. plurianulatum (Hedgcock) H. & P. Sydow, O. stenoreras (Robak) Melin & Nannf., and O. europhioides (Wright & Cain) Solheim] could not be unambiguously aligned with the ITS sequences of the O. piceae complex, so only members of the O. piceae complex were included in analyses. Of the species outside of the O. piceae complex, the ITS sequence of O. ips (Rumbold) Nannf. (AF198244) was reasonably similar, and O. ips was used as an outgroup taxon.

Two major clades in the O. piceae complex were inferred from parsimony analysis of the ITS sequences (Fig. 1). Ophiostoma piceae s.s., O. canum, O. floccosum and O. setosum formed a moderately supported group of species that are primarily found on conifer (Pinaceae) wood. Ophiostoma querci, O. catonianum, O. himal-ulmi, O. ulmi and O. novo-ulmi were found in a strongly supported branch of primarily hardwood (angiosperm) inhabiting fungi. The inferred hardwood-inhabiting clade was also seen in neighbor-joining analysis (Swofford 1998), as was the branch containing O. himal-ulmi and O. querci, but the conifer-inhabiting species did not resolve as a single clade.

In most cases, the well supported branches in the phylogenetic analysis (Fig. 1) were of isolates with unique cultural or morphological characteristics (Table II). The ITS sequences of O. piceae and O. canum were identical, though all of the examined O. piceae isolates had cylindrical to obovoid conidia, and the two examined isolates of O. canum had globose conidia at the tip of synnemata (Fig. 13). The synnema stipes of O. piceae isolates were consistently black, while some synnemata of O. canum had brown stipes. Ophiostoma floccosum formed red-brown synnemata, frequently with lateral knobs (Fig. 16). Another branch in the ITS tree contained isolates of the recently describe O. setosum (Uzunovic et al 2000), which is characterized by darkly pigmented synnemata with the vertical cells of the stipe extending into the conidial mass, forming a cup-shaped structure (Figs. 21–23).

As expected, the Dutch elm disease fungi O. ulmi and O. novo-ulmi were grouped as closely-related sister species (Fig. 1). The O. himal-ulmi ITS sequence was closer to that of O. querci than to that of O. ulmi. The O. querci clade included the only available isolate of O. catonianum and anomalous isolates from New Zealand and Viet Nam. Ophiostoma fagi had an ITS sequence identical to O. querci and is considered a synonym.

PCR-RFLP diagnostics.—Based on the ITS sequences used in Fig. 1, we identified restriction enzymes that would differentiate among the species in the O. piceae complex, except that no attempt was made to differentiate among the three Dutch elm disease species. Most species could be differentiated based on HaeII digestions (Table IV, Fig. 49). The HaeII restriction pattern for O. ulmi, O. novo-ulmi and O. himal-ulmi is the same, but it is distinct from that of O. querci and the other species (Table IV, Fig. 50). Ophiostoma piceae and O. setosum have the same re-
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<td>C969</td>
<td>AF198238</td>
<td><em>Quercus</em></td>
<td>United Kingdom</td>
<td>B</td>
<td>6.3</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C970</td>
<td>AF198239</td>
<td><em>Quercus</em></td>
<td>United Kingdom</td>
<td>A</td>
<td>6.8</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C984</td>
<td>2NZ-26</td>
<td><em>Pinus</em></td>
<td>New Zealand</td>
<td>A</td>
<td>6.8</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1085</td>
<td>AF198237</td>
<td><em>Fagus</em></td>
<td>Germany</td>
<td>A</td>
<td>5.6</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1159</td>
<td>CBS 102354</td>
<td><em>Pseudotsuga</em></td>
<td>Washington, USA</td>
<td>mixed</td>
<td>8.9</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1283</td>
<td>CBS 277.54, ATCC 24586, JCM 9745/Kåårik (as <em>Graphium aureum</em>)</td>
<td>Pinus</td>
<td>Sweden</td>
<td>B</td>
<td>6.3</td>
<td></td>
</tr>
<tr>
<td><em>Ophiostoma catonianum</em></td>
<td>C1084</td>
<td>AF198243</td>
<td><em>Pyrus</em></td>
<td>Italy</td>
<td>?</td>
<td>&lt;1</td>
<td></td>
</tr>
<tr>
<td><em>Pesotum</em> sp., near <em>O. querci</em></td>
<td>C1203</td>
<td>18NZ-128</td>
<td><em>Pinus</em></td>
<td>New Zealand</td>
<td>?</td>
<td>4.4</td>
<td></td>
</tr>
<tr>
<td><em>Pesotum</em> sp., near <em>O. querci</em></td>
<td>C1214</td>
<td>144-E/Blanchette</td>
<td><em>Aquilaria crassa</em></td>
<td>Viet Nam</td>
<td>?</td>
<td>12.4</td>
<td></td>
</tr>
<tr>
<td><em>Pesotum</em> sp., near <em>O. querci</em></td>
<td>C1215</td>
<td>AF198240</td>
<td><em>Nothofagus</em></td>
<td>New Zealand</td>
<td>?</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td><em>Ophiostoma ulmi</em></td>
<td>C55</td>
<td>UM C-20/Blanchette</td>
<td><em>Ulmus</em></td>
<td>Minnesota, USA</td>
<td>?</td>
<td>17.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1089</td>
<td>CBS 137.36 (as <em>O. ips</em>)</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>21.2</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1182</td>
<td>AF198232</td>
<td><em>Ulmus</em></td>
<td>Netherlands</td>
<td>?</td>
<td>13.3</td>
<td></td>
</tr>
<tr>
<td><em>Ophiostoma novo-ulmi</em></td>
<td>C1186</td>
<td>CBS 296.87, WCS 811/Elgersma</td>
<td><em>Ulmus</em></td>
<td>Netherlands</td>
<td>?</td>
<td>13.8</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C48</td>
<td>UM C56, Aggressive W-Z/Blanchette</td>
<td><em>Ulmus</em></td>
<td>United Kingdom</td>
<td>?</td>
<td>10.3</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C510</td>
<td>AF198236</td>
<td><em>Ulmus</em></td>
<td>Iowa, USA</td>
<td>?</td>
<td>&lt;1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C913</td>
<td>Jacobs</td>
<td><em>Ulmus</em></td>
<td>Illinois, USA</td>
<td>?</td>
<td>9.1</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C943</td>
<td>Jacobs</td>
<td><em>Ulmus</em></td>
<td>Illinois, USA</td>
<td>?</td>
<td>13.5</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1184</td>
<td>CBS 377.79/Meulemans</td>
<td><em>Ulmus</em></td>
<td>Belgium</td>
<td>?</td>
<td>13.5</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1185</td>
<td>AF198235</td>
<td><em>Ulmus</em></td>
<td>Russia</td>
<td>?</td>
<td>11.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1187</td>
<td>CBS 297.87, WCS 802/Scheffer</td>
<td><em>Ulmus</em></td>
<td>Netherlands</td>
<td>?</td>
<td>&lt;1.0</td>
<td></td>
</tr>
<tr>
<td><em>Ophiostoma himal-ulmi</em></td>
<td>C1183</td>
<td>AF198233</td>
<td><em>Ulmus</em></td>
<td>India</td>
<td>?</td>
<td>1.4</td>
<td></td>
</tr>
<tr>
<td></td>
<td>C1306</td>
<td>AF198234</td>
<td><em>Ulmus</em></td>
<td>India</td>
<td>?</td>
<td>3.5</td>
<td></td>
</tr>
</tbody>
</table>

<sup>a</sup> Isolate numbers from the collection of the senior author.

<sup>b</sup> Designations of mating type as either A/B or ± were based on compatibilities with tester strains provided by C. Brasier, A. Uzunovic, or J. Webber, using their nomenclature. Isolates producing perithecia and ascospores without pairings ("mixed") are mixtures (heterokaryons or mixtures of mycelia) of both mating types.

<sup>c</sup> Extent of radial growth on MEA after 7 d at 32 °C.
Fig. 1. One of four most parsimonious trees of the *Ophiostoma piceae* complex based on 616 characters, including gaps, of the ITS-1, 5.8S, and ITS-2 regions of the rDNA operon. The tree is rooted to *Ophiostoma ips*. Branches with strong support (≥ 75%) are bolder, with bootstrap values ≥ 50% indicated above the branches. Total tree length = 181 steps, consistency index = 0.8122, retention index = 0.9663. Asterisk indicates a culture from the holotype.
Table II. Distinguishing characteristics for species in the *Ophiostoma piceae* complex

<table>
<thead>
<tr>
<th>Species</th>
<th>Knobs on Cupulate Synema</th>
<th>Synema synema conidial mass</th>
<th>Shape of synema stipe</th>
<th>Color of synema stipe</th>
<th>Mycelium with concentric rings</th>
<th>Culture aroma *</th>
<th>Growth (mm) at 32°C</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>O. piceae</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>0</td>
</tr>
<tr>
<td><em>O. canum</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
<tr>
<td><em>O. floccosum</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
<tr>
<td><em>O. querci</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
<tr>
<td><em>O. setosum</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
<tr>
<td><em>O. atomophilum</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
<tr>
<td><em>O. nana-philum</em></td>
<td>No</td>
<td>White</td>
<td>Cylindrical to Obvoid</td>
<td>Dark-brown</td>
<td>Rare</td>
<td>None</td>
<td>1-5</td>
</tr>
</tbody>
</table>

Species of the *Ophiostoma piceae* complex.—Based on ITS sequence analysis, mating reactions, tests for growth at 32°C, and morphology, the following species are recognized in the complex.


Commentary. Lectotype (BPI 595978 = FP 14395) and other material (BPI 595979, BPI 595977) from Picea abies sapwood in Germany, collected by Münch, was deposited in BPI, apparently by Hedgcock. Although there is some mold growth on these specimens, the lectotype has, in addition to synnemata and perithecia, the Sporothrix synanamorph (with prominent denticles) typical of the O. piceae complex. The culture from the type (CBS 108.21 = C1087) forms synnemata on wood and is of the B mating type (TABLE I). This species is most commonly found on members of the Pinaceae throughout Europe, North America, and Japan (Brasier and Kirk 1993, Halmschlager et al 1994, Kim et al 1999, Morelet 1992, Pipe et al 1995). We have also identified isolates of O. piceae on the exotic Pinus radiata from New Zealand (TABLE I) and Chile.

Both the teleomorph and anamorphs of this spe-
cies lack distinctive features, which has led to the proposal of numerous synonyms, all of which we have excluded from *O. piceae*. Synnemata stipe lengths of *O. piceae* and *O. quereri* overlap, but those of *O. piceae* are larger (Hamschlager et al 1994). The size of the ascospores (2.5–4.5 × 1.2–2 μm) and range in size of synnematosus conidia (5–15 × 1–3.5 μm) (Upadhyay 1981) of *O. piceae* overlap with the sizes of most other species in the complex. However, ITS sequences (Fig. 1, Kim et al 1999), mating reactions (Table I, Brasier and Kirk 1993), dark protoperithecia, the inability to grow at 32 C (Table II, Brasier and Stephens 1993), and absence of a distinctive aroma or concentric rings on MEA (Table II) distinguish isolates of *O. piceae* from the more common and widespread *O. quereri*. The synnemata of *O. piceae* are also similar to those of *O. setosum*, but the cup-like, conidial-bearing structure at the top of the synnemata of *O. setosum* distinguishes this species from the rest of the *O. piceae* complex. Red-brown synnemata distinguish *O. floccosum* from *O. piceae* and other members of this complex. Although *O. canum* has the same ITS sequence as *O. piceae*, the former is distinguished by its globose conidia and brown synnematos stipe.


Commentary. The lectotype of *O. canum* (BPI595767 = BPI 2107) and two other specimens (BPI 595765 = BPI 2110 and BPI 595766 = FP 14394) from Münch were deposited in BPI by Hedgcock. The lectotype has perithecia and ascospores, while the other two specimens have synnemata and a *Sporothrix* anamorph but no perithecia. In addition to the isolates listed in Table I, we have examined Norwegian isolates NFRI 1652/2 (C1478) and NFRI 97-33/47 (C1477), both from *Tomicus minor* Hartig or stained *Pinus sylvestris* sapwood around galleries of this beetle, and globose conidia are also formed by these isolates. Mathiesen (1950, Mathiesen-Käärik 1960) noted the association of *O. canum* with *T. minor*, and this beetle association distinguishes *O. canum* ecologically from *O. piceae*. Mathiesen-Käärik (1960) also noted physiological differences between *O. canum* and *O. piceae*. To date, *O. canum* is known only from pine in Europe, from Scandinavia and Germany.

Mathiesen (1950) and Upadhyay (1981) noted ascospores 5–6 × 1.5–2.5 μm and 4–6.5 × 1.5–3 μm, respectively, in *O. canum*, somewhat larger than those in *O. piceae*. The globose conidia of *O. canum* are particularly distinctive (Fig. 13) and separate it from all other members of the complex. Although we found that the ITS sequence of *O. canum* is the same as that of *O. piceae*, Kim et al (1999) reported that PCR primers specific for the ITS region of *O. piceae* were unable to amplify this fragment in *O. canum*. Hauser et al (1993) found that 259 of 261 aligned bases of the 26S rDNA gene were identical between a culture from the holotype of *O. piceae* and a Norwegian isolate of *O. canum*.

We have not been able to produce perithecia or ascospores with the two isolates (Table I) of *O. canum* that we have studied closely, perhaps because these two isolates are of the same mating type. However, we were able to produce a few perithecia when either of these two isolates were paired with mating type A testers of *O. piceae*. Ascospores produced from these hybrid crosses were generally inviable, but viable ascospore progeny were recovered from one perithecium of the cross between isolate C967 of *O. piceae* and isolate C114 of *O. canum*. Ten viable progeny were recovered, with globose conidia of *O. canum* and the elliptical conidia of *O. piceae* seen among the progeny. The fact that the *O. canum* isolates are only partially interfertile with *O. piceae* strains suggests that there is an intersterility barrier between these two species and further supports recognition of *O. canum* as distinct (Brasier 1993, Harrington and McNew 1998).


Anamorph. *Pesotum aureum* (Hedgcock) McNew et Harrington, comb. nov.


Commentary. Although well described by Mathiesen (1951), this species has been rarely reported by others, probably because it was thought to be a morphological variant of *O. piceae*. Also, perithecia and ascospores are not commonly seen in cultures of *O. floccosum*, and perithecia often take many weeks to form in mating tests. We have examined specimens and cultures of this species from Europe, North America, Australia, and New Zealand (Table I), and also isolates from Korea. All of these isolates were from Pinaceae.
### Table III. *Ophiostoma* species with *Pesotum*-like anamorphs but without a *Sporothrix* synanamorph

<table>
<thead>
<tr>
<th>Teleomorph</th>
<th>Anamorph</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>O. clavatum</em> Mathiesen</td>
<td>Pesotum-like</td>
<td>Hunt (1956), Mathiesen (1951)</td>
</tr>
<tr>
<td><em>O. clavigerum</em> (Robinson-Jeffrey &amp; Davidson) Harrington</td>
<td><em>Graphiocladiella clavigerum</em> Upadhyay = <em>Pesotum clavigerum</em> Okada &amp; Seifert</td>
<td>Okada et al. (1998), Robinson-Jeffrey and Davidson (1968), Upadhyay (1981)</td>
</tr>
<tr>
<td><em>O. obscura</em> (Davidson) von Arx</td>
<td>Pesotum-like</td>
<td>Davidson (1935), Hunt (1956)</td>
</tr>
</tbody>
</table>
No holotype for *O. floccosum* was designated by Mathiesen, but herbarium material is apparently available in Statens Skogsforskningsinstitut Experimentalfältet, Sweden (Hunt 1956). A culture from Käärik deposited in CBS (799.73) is considered authentic for the type but does not produce perithecia and ascospores, unless mated with – (minus) mating type strains. Tester strains of an intersterility group identified by J. Webber proved to be *O. floccosum* in morphology and ITS sequence. We deposited a dried specimen (BPI 746439) with perithecia and ascospores that resulted from pairing CBS 799.73 (from the holotype) and C1013 (a tester strain from Britain provided by Webber).

Mathiesen (1951) noted the relatively short ostiolar hyphae of this species when compared to *O. piceae*. We see little difference between the two species (Figs. 5, 15), though ascospore masses at the top of perithecia are relatively small for *O. floccosum*. Synnemata of *O. floccosum* are distinctly red-brown, and lateral knobs on synnemata (Fig. 16) are common in all of the cultures of *O. floccosum* that we have examined. Also, secondary synnemata are frequently found emanating from the conidial mass on top of primary synnemata. Conidial masses on synnemata are yellow compared to white in fresh spore masses of other species in the complex.

Hedgcock (1906) described *Graphium aureum* from stained sapwood of *Pinus strobus* in Wisconsin. His description includes white to yellow conidial masses and stipes that are white at first, changing to yellow and often to dark brown at the base. The yellow conidial masses described by Hedgcock (1906) are typical for *O. floccosum*, though our cultures generally produce red-brown stipes instead of yellow to brown stipes. Hedgcock’s type material is available (BPI 448701 and BPI 448702), and both specimens contain red-brown synnemata and black protoperithecia, which are also common for *O. floccosum*. A Sporothrix-like anamorph was noted by Hedgcock and was observed in the BPI specimens. Thus, we believe that Hedgcock’s *Graphium* species is the anamorph of *O. floccosum*, and we have transferred his fungus to *Pesotum*. A culture deposited by Käärik (CBS 277.54 = ATCC 24586 = JCM 9745 = C1283) as *G. aureum* mated with *O. queri* tester strains as discussed above and does not represent *G. aureum*.


**Anamorph.** *Pesotum cupulatum* McNew et Harrington sp. nov. Figs. 21-26

Coloniae olivaceae-brunneae de infra, in agarō extractī mali diam. 55 mm attingēns post 11 dies ad 25 C, odore

dulci. Incremento nullo ad 32 C. Synanamorpha primaria stipite 80–820 μm longo, cum cellulis fuscae protrudens in zonam conidiogenam, calyx formans. Conidia unicellularia hyalina, cylindrica vel obovoidea, 3.0–5.5 μm × 1.0–2.0 μm. Synanamorpha altera, simili *Sporotrichi* in denticules conspicui ubi sporae portatae et quoque in catena acropeta conidia holoblastica efferens. 

Colonies on malt extract agar at 25 C attaining a diameter of 55 mm in 11 d, appearing olive-green to gray from above, olive-green to brown from below, with concentric rings developing on the upper surface of the mycelium with age, with a distinctly sweet aroma. Cultures not inhibited by cycloheximide. No growth at 32 C on malt extract agar. *Hyphae* mostly submerged, aerial hyphae appressed, hyaline, smooth-walled, 1.0–3.0 μm wide, to brown, smooth or rough-walled, 2.5–4.0 μm wide. Conidiophores of two types: *synnemata* (Figs. 21–23) single or in groups of two to three, simple or rarely branched, macroematous. Stipe dark brown to black at base, becoming lighter towards apex, 80–820 μm (mean 290 μm) long, base 10–40 μm (mean 25 μm) wide, tapering to 6–32 μm (mean 16 μm) near the apex and flaring to 14–120 μm (mean 45 μm). Many of the synnema stipe cells extend into the mass of conidiogenous cells and conidia, forming a cup of brown to black, setae-like structures. Conidiogenous cells proliferating sympodially with inconspicuous conidial scars. *Synnematous conidia* (Fig. 26) hyaline, one-
TABLE IV. Restriction fragments of the amplified ITS product from species in the Ophiostoma piceae complex

<table>
<thead>
<tr>
<th>Species</th>
<th><em>HaeIII</em> fragments (bp)</th>
<th><em>DdeI</em> fragments (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>O. piceae/O. canum</em></td>
<td>400, 200, 120</td>
<td>290, 270, 190</td>
</tr>
<tr>
<td><em>O. floccosum</em></td>
<td>280, 200, (120)^a</td>
<td>290, 270, 190</td>
</tr>
<tr>
<td><em>O. setosum</em></td>
<td>400, 200, 120</td>
<td>270, (190)^a, 100</td>
</tr>
<tr>
<td><em>O. querci/O. catanorum</em></td>
<td>520, 200</td>
<td>290, 270, 190</td>
</tr>
<tr>
<td><em>O. ulmi/O. novo-ulmi/</em></td>
<td>520, 170</td>
<td>290, 270, 190</td>
</tr>
</tbody>
</table>

^a Band sizes in parentheses are for two co-migrating bands.

celled, mostly guttulate, cylindrical to obovoid with rounded apices and slightly tapering bases, which may be truncate to rounded, 3.0–5.5 μm (mean 4.0 μm) long and 1.0–2.0 μm (mean 1.6 μm) wide, accumulating in a white, gelatinous mass at the apex of the synnema. Micronematous conidiophores typical of the genus *Sporothrix* (FIGS. 24, 25), arising terminally or laterally from hyphae, 11–156 μm (mean 73 μm) long, 1.0–4.0 μm wide at the base, tapering slightly at the apex, sometimes septic. Conidia produced at the apex of the conidiogenous cell (FIGS. 24, 25), which has prominent denticles, 0.5–1.5 μm long, where conidia have abscised along a rachis 1.5–21 μm (mean 5.5 μm) long. Conidiogenous cells may proliferate at or below the rachis. *Sporothrix conidia* hyaline, smooth-walled, 6.5–24 μm (mean 12.0 μm) long, 2.0–3.5 μm (mean 2.5 μm) wide, with a rounded apex and tapering base, often giving rise to secondary conidia at the apex and having denticles at the point of dehiscence of the secondary conidia. Secondary *conidia* hyaline, smooth-walled, guttulate, cylindrical to obovoid, 3.0–8.0 μm (mean 5.5 μm) long, 1.0–2.5 μm (mean 2.0 μm) wide, with a rounded apex and tapering at the base. No protoperitheciun or peritheciun seen.

**HOLOTYPE.** USA. WASHINGTON: Aberdeen, from stained wood of *Pseudotsuga menziesii*, Dec 1997, T. Harrington, BPI 746441, from isolate C1194 (CBS 102358).

**Etymology.** Refers to the cup-like structure at the top of the synnema.

**Commentary.** Cultures representing the two mating types of a new *Ophiostoma* species (*O. setosum*) from Canada were kindly provided to us by A. Uzunovic (Uzunovic et al 2000) after our initial mating studies were completed. Pairings of our *P. cupulatum* isolates with the tester strains of *O. setosum* showed that all of our isolates were of the same mating type (B, TABLE I). After repeated pairings at 20 C, the isolates of *P. cupulatum* from the USA and New Zealand (TABLE I) were able to mate with the tester strains provided by Uzunovic. Morphologically, our isolates and the Canadian isolates (Uzunovic et al 2000) are identical.

Synnemata of *O. setosum* are similar in size and pigmentation to those of *O. piceae* and *O. querci*, but the apex of the synnemata of *O. setosum* is unique in its cup-like structure formed by extension of the external stipe cells into the conidial mass (FIGS. 21–23). The concentric rings of aerial mycelium found in most isolates and the distinctively sweet aroma of cultures of *O. setosum* also distinguish this species from *O. piceae* (TABLE II). *Ophiostoma setosum* fails to grow at 32 C, as does *O. piceae*. In ITS sequence, *O. setosum* is close to *O. piceae*, and we have found *O. setosum* only on wood of conifers (Pinaceae), the most common substrate of *O. piceae*.

We originally isolated *O. setosum* from stained wood of *Pinus radiata* in New Zealand. Additional isolations were made from lumber of *Pseudotsuga menziesii* and *Tsuga* sp. from Washington, USA and from *Pinus radiata* wood from California. Uzunovic et al (2000) identified isolates from *Picea*, *Pinus*, and *Tsuga* in British Columbia, Canada and Oregon, USA. Thus, the species appears endemic to the Pacific Coast of North America on various Pinaceae and was likely introduced to New Zealand. Synnemata of this species are commonly found in association with *O. piceae*, *O. querci*, and *O. floccosum* on bluestained wood.

**Ophiostoma querci** (Georgévitch) Nannf., in Melin and Nannf., Sven. Skogsvarsfordens Tidskr. 32: 408. 1934.

**Figs.** 27–36


= *Ceratostomella fagi* Loos, Arch. Mikrobiol. 3: 376. 1932.


HARRINGTON ET AL: OPHIOSTOMA PICEAE


Commentary. Unfortunately, no type material or culture was preserved by Georgévitch, and no Latin description was made. However, this name has been commonly applied to the so-called “hardwood” form of the O. piceae complex, and we continue to apply the name in this sense. Morelet (1992) designated neotype material (PFN 1463) for O. querci, which was derived from mating two French isolates (CBS 366.93 and CBS 367.93). We have deposited a dried specimen (BPI 746440) with perithecia and ascospores resulting from a cross of two British isolates, C969 and C970.

Ophiostoma querci is difficult to distinguish morphologically from O. piceae, though ITS sequencing shows it to be only distantly related (Fig. 1). It is common to find small, glistening drops of ascospores along the neck of O. querci perithecia (Fig. 30),
which are due to exudations of ascospores as the perithecial neck elongates. We have not seen this in perithecia of the other species in the *O. piceae* complex, but we have seen such ascospore droplets along the neck of *O. pluriannulatum* perithecia. Most isolates of *O. querci* and *O. piceae* form protoperithecia on MEA, but those of *O. querci* are a light, golden-brown color (Figs. 27, 28), while those of *O. piceae* are black (Figs. 2, 3). Most isolates of *O. querci* have a nut-like aroma when growing on MEA, in contrast to an indistinct aroma in cultures of *O. piceae* (Table II). Also, *O. querci* isolates grow at 32°C, while *O. piceae* isolates do not (Brasier and Stephens 1993, Table I). Many isolates of *O. querci* form concentric rings of aerial mycelium on MEA, similar to isolates of the Dutch elm disease fungi (Table II).

In pairings, *O. querci* produces perithecia and ascospores when isolates of opposite mating types are paired. An isolate (CBS 236.32) of *O. fagi* from the holotype had the ITS sequence of *O. querci* and matched with this species. It is, therefore, considered a synonym of *O. querci*, as was suggested by Brasier (1993). Three other *Ophiostoma* species (*O. robinis*, *O. kubanicum*, and *O. valachicum*) from *Quercus* in eastern Europe are likely synonyms of *O. querci* (Przybyl and de Hoog 1989), but there appears to be no authentic material available of these three species (Przybyl and de Hoog 1989). From descriptions by Potlajczuk and Sekunova (1985), *O. robinis*, *O. kubanicum*, and *O. valachicum* are indistinguishable from *O. querci*. Both *O. robinis* and *O. kubanicum* have been isolated from *Quercus* from central Europe to Azerbaijan (Brasier and Kirk 1993). An isolate of *O. robinis* from Azerbaijan by Guseinov paired with testers of *O. querci* to produce perithecia and ascospores (Table I), as was reported by Brasier and Kirk (1993).

Both Morelet (1992) and Okada et al (1998) list *Pesotum pirinum* as the anamorph of *O. querci*. We consider *O. catania* and its anamorph, *P. pirinum*, as distinct from *O. querci* and *O. piceae*, leaving no clear anamorph name for *O. querci*. An isolate deposited as *Graphium aureum* (CBS 277.54, C1283) had the ITS sequence of *O. querci*, and it paired with our tester strains of *O. querci*. However, examinations of the holotype specimen and description of *G. aureum* (Hedgcock 1906) show that it is the anamorph of *O. floccosum*, as noted above, and CBS 277.54 was misidentified as *G. aureum*. Should *O. robinis* prove to be a validly described synonym of *O. querci*, then the anamorph name *Graphium robinis* is available as a basionym for the anamorph of *O. querci*.

*Ophiostoma querci* appears to be a widespread species and is commonly found on conifers and hardwoods throughout Europe and North America (Brasier and Kirk 1993, Halschlaghler et al 1994, Kim et al 1999, Morelet 1992, Pipe et al 1995). In addition to isolates from South Africa and New Zealand listed in Table I, we have identified isolates of *O. querci* from *P. radiata* in Australia, *Eucalyptus* sp. in Uruguay and *Pinus* sp. in Korea. In Europe, it apparently occurs more frequently on hardwoods, especially oak, than on conifers (Brasier and Kirk 1993, Halschlaghler et al 1994, Morelet 1992, Pipe et al 1995). In North America and the Southern Hemisphere, however, sapwood of Pinaceae is a common substrate.


Commentary. Although there appears to be no type specimen for *O. catania* (Hunt 1956), a culture of *O. catania* deposited by Goidanich is available from CBS (263.35). The ITS sequence of this isolate is unique but near that of *O. querci* (Fig. 1). The culture does not form protoperithecia, perithecia, or synnemata, and pairings with testers of the other species in the complex failed to result in perithecia. A *Sporothrix* anamorph was found in the culture, however. This culture grows very slowly at 32°C, in contrast to *O. querci* (Table I).

Goidanich (1935) described this species from *Pyrus* (pear) in Italy. His description of *O. catania* indicates that the perithecial necks are shorter and the ostiolar hyphae longer than those of *O. piceae*, *O. querci*, and *O. floccosum*. He also states that single ascospore and single conidial isolates produce perithecia and ascospores. Thus, *O. catania* is homothalic, in contrast to the heterothallic mating system of all other species in the complex.


Synnematous Ophiostoma species without Sporothrix synanamorphs.—A number of Ophiostoma species have been noted to have synnematous anamorphs but no Sporothrix synanamorph (TABLE III). Ophiostoma novo-ulmi (Hutchison & Reid) Rulamort was said to rarely form synnema (Hutchison and Reid 1988a), but we have not seen synnema in a dried culture from the holotype [WIN (M) 869] or in living cultures (UAMH 9556-9) from New Zealand (Hutchison and Reid 1988a). The original description may have been from a culture of more than one species. Perithecia and ascospores as described by Hutchison and Reid (1988a) and as seen in the holotype look like those of O. plurianulatum, which is not reported to have synnema (Hunt 1956, Upadhyay 1981).

Reports of synnemata in Ophiostoma species other than those in the O. piceae complex have been of species without known Sporothrix synanamorphs, though many produce Hyalorhinocladiella synanamorphs. These species (TABLE III) are excluded from the O. piceae complex. Two such species are discussed below.


Commentary. An isolate deposited as Graphium fragrans (C1224 = CBS 279.54 = ATCC 24590) by Käärik likely represents the species. However, this Swedish isolate no longer produces synnema but does produce Leptographium-like or Hyalorhinocladiella-like conidiophores (Figs. 37–39) that may represent degenerate synnemata. No denticles typical of Sporothrix are seen in the culture. The conidia (Fig. 40) of this species are longer than those of members of the O. piceae complex.

This species does appear to be an anamorph of an Ophiostoma species based on ITS sequence (AF198248), but it does not fall into the O. piceae complex (data not shown). Similar cultures with almost identical ITS sequences were isolated from pine wood in California, New Zealand and Australia (unpub).

Okada et al (1998) list CBS 219.83 as authentic for the type of G. fragrans, but this isolate was collected by Solheim, not Käärik, and CBS 279.54 should be authentic for the type. Okada et al (1998) found sympodial proliferation of the conidiogenous cells of synnematosus and mononematous conidiophores of CBS 219.83, and they also found that the 18S rDNA sequence of this isolate was similar to that of other Ophiostoma species.

**Commentary.** The ITS sequence of *O. cucullatum* (AF198246, from isolate C1216 = NFRI 81–83/2, collected by Solheim) differed in only a single base substitution from that of *Graphium erubescens* (CBS 278.54 = JCM 9747 = C1222, from the holotype) and another culture deposited by Kåårik as *G. album* (CBS 276.54 = JCM 9744 = C1225, ITS sequence = AF198247). This isolate of *G. album* is morphologically identical to *G. erubescens* and is distinct from specimens representing *G. album* as emended by Hedgcock (1996). We believe that Kåårik’s isolate of *G. album* was misidentified and that *G. erubescens* is the anamorph of *O. cucullatum*.

The ITS sequences of *O. cucullatum* and *G. erubescens* were not similar to those of other members of the *O. piceae* complex. Okada et al (1998) found the 18S rDNA sequence of *O. cucullatum* to be similar to that of *O. europhioides*, which has a *Leptographium* anamorph. Likewise, Hausner et al (1993) found the partial 28S rDNA sequence of *O. cucullatum* to be similar to those of other *Ophiostoma* species with *Phialographium* and *Leptographium* anamorphs.

All three cultures of *O. cucullatum* (C1216, C1222, C1225) failed to form *Sporothrix* conidiophores on MEA but did form synnemata with loosely-to-tightly fused outer stipe cells (Figs. 41–43), with gradations to mononematous (*Leptographium*-type) conidiophores (Fig. 45). In general, the aerial synnemata are more tightly woven, and the synnemata submerged in the agar medium are a loose collection of *Leptographium*-like conidiophores. The conidiogenous cells on top of the synnematus and mononematous conidiophores are phialidic based on light microscopy (Figs. 43, 45), as described for the synnemata of *O. cucullatum* (Solheim 1986). Okada et al (2000) reported that the conidiogenous cells of *P. erubescens* (isolate CBS 278.54) usually proliferate percurrently but also have an intermediate mode between sympodial proliferation and phialidic ontogeny based on scanning electron microscopy.

The anamorph of *S. sagmatosporae* also has conidiogenous cells that are phialidic in light microscopy (Seifert and Okada 1993), and this anamorph was the basis for the synnema-forming genus *Phialographium* Upadhyay & Kendrick. Okada et al (1998) consider the synnematus anamorphs of *O. sagmatosporae* and *O. cucullatum* to be *Pestotum* species, and they (Okada et al 2000) formally transferred *G. erubescens* to *Pestotum*. However, the phialidic conidiogenous cells, the loose aggregation of *Leptographium*-like conidiophores, and the absence of a *Sporothrix* anamorph lead us to believe that these species are distinct from *Pestotum* as originally conceived by Crane and Schoknecht (1973). Further, *O. cucullatum* and *O. sagmatosporae*, like most other *Ophiostoma* species with *Leptographium* or *Phialographium* anamorphs, have ascospores with thickened outer walls (sheaths) that are cucullate, to rectangular, to triangular, depending on the view (Solheim 1986). In contrast, ascospores in the *O. piceae* complex are allantoid to orange section shaped, and are relatively thin-walled, with no visible sheath.

*Graphium* species excluded from *Ophiostoma.*—There are numerous *Graphium*-like species that are related to ascomycetes other than *Ophiostoma* (Okada et al 1998), and it is beyond the scope of the present paper to discuss these in detail. However, we did examine material of five *Graphium* species from Hedgcock’s specimens in BPI and other CBS cultures, and these examinations suggest that they are not closely related to *Ophiostoma* species.

= *Ceratopodium album* Corda, l.c. Fung. 1: 20. 1837.

**Commentary.** No type material of Corda’s specimen is available (Seifert, pers comm), and his illustrations give little indication of the affinities of this species. It was found on *Betula* in Bohemia and described by Saccardo (1886) as having conidial masses that were white. Hedgcock (1906) emended the species description to include conidial masses white to creamy-yellow, or light brown when old and dry, and our examination this specimen (BPI 448665, from Fagus in Arkansas, USA) showed the conidial masses to be yellow-gold. The yellow-gold to light-brown conidial masses of the BPI specimen suggests that *G. album* as emended by Hedgcock is not an *Ophiostoma* species. However, Hedgcock’s fungus may not be *G. album*, and the disposition of Corda’s species remains in question.

A culture from *Pinus* in CBS (276.54, deposited by Kåårik) is listed as *G. album*, but the culture does not match the BPI specimen or Hedgcock’s emended description of *G. album*. Rather, Kåårik’s culture is conspecific with her *Graphium erubescens* (Mathiesen-Kåårik 1953). As discussed earlier, CBS 276.54 has the ITS sequence of *O. cucullatum* (anamorph = *Phialographium erubescens*). As Kåårik’s culture is from
pine, it seems unlikely that it is *G. album* as described by Corda.


**Commentary.** The holotype specimen (BPI 448676) and co-type (BPI 448675) are in poor condition, but both have synnemata with dark stipes and black conidial masses. The species was described from galleries of ambrosia beetles in *Pinus arizonica* in Arizona, USA. It is possible that Hedgcock was working with a mixture of an *Ambrosiella* species with a *Graphium* species. The black conidial masses on the synnema of the *G. ambrosiigerum* specimens are not typical for anamorphs of *Ophiostoma* species.


**Commentary.** Seifert (1985) suggested that the name *G. rigidum* had been frequently used for fungi with black synnemata, and he considered the species a *nomen dubium*. Siemaszko (1939) considered *G. rigidum* as a synonym of *G. penicillioides*, and *G. rigidum* may be a member of the *G. penicillioides* complex (Okada et al. 1998). A specimen (BPI 448820) from *Quercus rubra* sapwood from Indiana, USA deposited by Hedgcock in BPI as the emended type of *G. rigidum*, may be of more than one fungus. The synnemata in the specimen have black stipes from the base up to the yellow-gold conidial masses.

*Graphium rubrum* Rumbold, Phytopathology 24: 300. 1934.

**Commentary.** A specimen deposited by Rumbold (BPI 448830) can serve as the holotype of *G. rubrum*, and it shows black synnematous stipes with pinkish-red to yellowish-red conidial masses, and a few synnemata have gray conidial masses. A culture from the holotype (CBS 210.34) is no longer producing synnemata but is producing phialide-like structures on short, simple, hyaline conidiophores (Figs. 46–47). Okada et al. (2000) also observed phialides in CBS 210.34 using scanning electron microscopy.

Rumbold (1934) reported *G. rubrum* from *Quercus, Populus, Liquidambar* and *Pinus* in the USA. Goidanich (1936) described *G. silanum* from *Pinus* in Italy, and a culture (CBS 206.37) deposited by him produces no synnemata but does produce phialide-like structures on short, simple, hyaline conidiophores (Fig. 48), similar to those of *G. rubrum*. Also, CBS 206.37 has an ITS sequence identical to that of Rumbold’s isolate of *G. rubrum*. It is possible, however, that CBS 206.37 is not *G. silanum*, as Goidanich described *G. silanum* as having a micromatous state with prominent denticles and ramoconidia, and it seems unlikely that it is *G. album* as described by Corda.

![Fig. 49. Ethidium bromide stained agarose (2%) gel of HaeII and DdeI digestion products of the amplified ITS rDNA region from isolates of *Ophiostoma setosum* (Os), *Ophiostoma floccosum* (Of), *O. piceae* (Op), and *O. querci* (Oq). Co-migrating bands are indicated by arrows. One-hundred base pair ladders (M) are on either side of the gel.](image-url)
CBS 206.37 does not form such a Sporothrix-like anamorph.

The ITS sequence (AF198245) of Rumbold’s isolate CBS 210.34 did not match well with our sequences of Ophiostoma species. A BLAST 2.0 (NCBI) search showed this sequence to be closest to Phialophora gregata (Allington and Chamberlain) Gams, which may be an anamorph of an unknown discomycete (Paulin and Harrington 2000). The 18S rDNA sequence of Rumbold’s culture also places it among the discomycetes (Okada et al 2000).


Commentary. Seifert (1985) examined what was considered authentic material of this species and described the green synnemata and acroleurogenous phialides. Hedgcock (1906) noted green synnemata stipes and gray or green conidial masses, and we also found very dark conidial masses on his specimen (BPI 448617). Based on Seifert’s description and Hedgcock’s specimen, G. smaragdimum is not a likely member of the O. piceae complex. This species may have an unnamed Claussenomyces (discomycete) teleomorph (Okada et al 2000).

DISCUSSION

Although there have been earlier studies distinguishing the so-called conifer form of O. piceae from the so-called hardwood form, and the Dutch elm disease fungi have been clearly delimitated from each other, this is the first comprehensive assessment of the entire complex. The ITS sequences of the nine recognized species were very similar to each other but quite distinct from the ITS sequences of O. ips, O. simplex, O. galeiformis, O. cucullatum, O. europhioides, O. pluriannullatum, O. coronatum, O. perfectum, O. nigrocarpum, O. stenoceras and other Ophiostoma species (data not shown). All members of the O. piceae complex form a Sporothrix synanamorph with prominent denticles in addition to the Pesotum synanamorph, and the presence of both synanamorphs sets the complex apart from the rest of Ophiostoma. Thus, this unique combination of anamorphs and the ITS sequence analysis suggest that O. piceae complex is a monophyletic group.

Graphium-like fungi outside the O. piceae complex.—Synnemata are efficient fruiting bodies for placing conidia in the path of small animals for acquisition and dispersal, and it is not surprising that many evolutionary lines of fungi have converged on such conidiomata. In a recent review of ascomyete-forming fungi with Graphium-like synnemata, Okada et al (1998) found that such synnemata are formed by species of the Microcales (pyrenomycetes) and Chaetothyriales (loculoascomycetes), and they recommend that the genus name Graphium be used only for the species with microascalen affinities, such as G. penicillioides. Further, we found that the ITS sequence of G. rubrum is closer to Phialophora gregata and discomycetes than to pyrenomycetes or loculoascomycetes, and G. rubrum produces a phialidic micronematous state, as does the synnematous Dendrostilbella smaragdina, which also appears to be a discomycete anamorph (Okada et al 2000). Species in Ophiostoma tolerate cycloheximide at high concentrations (Harrington 1981), and this can aid in separating synnema-forming species with Ophiostoma affinities from those of other ascomycetes. However, there are other ascomycetes, including some true Graphium species, like G. penicillioides, that are not related to Ophiostoma but do tolerate cycloheximide.

Even within Ophiostoma, synnemata may have evolved more than once. Ophiostoma is a large genus with a wide variety of anamorphs, and many species have more than one synanamorph. In addition to the Leptographium-type (mononematous, with a stipe and a penicillately branched conidiogenous apparatus) and the Graphium-type (synnematous), some species of Ophiostoma, such as the anamorph of O. clavigerum, show both types of conidiomata. Graphiocladiella was proposed by Upadhyay (1981) to accommodate anamorphic fungi with both mononematous and syn-
nematous conidiomata. Harrington (1988) suggested that such intermediate types could be better accommodated in Graphium (Pesotum) than in Leptographium, but we now suspect that many such species have greater affinity with Leptographium.

Variability in conidiogenesis and stipe configuration among the anamorphs of Ophiostoma has led to a multitude of anamorph genera and continued controversy (Harrington 1988, Seifert and Okada 1993, Wingfield et al 1991). In some Ophiostoma species (TABLE III), the synnemata appear to be a loose aggregation of Leptographium conidiophores, without fused stipe cells. In our examination of Ophiostoma species, only members of the O. piceae complex have proven to produce both synnemata with fused stipe cells and a Sporothrix synanamorph (micronematous conidiophores with prominent denticles, Hoog 1974, 1993). Micronematous conidiophores are seen in many of the species listed in TABLE III, but these micronematous conidiophores lack prominent denticles at the conidiogenous cell apex and appear to be nearer to Hyalorhinocladiella (Hoog 1993) or to be degenerative Leptographium-type conidiophores. The conidiogenous cells of Pesotum and Leptographium resemble those of Hyalorhinocladiella, and it is possible that the macronematous conidiomata evolved from the Hyalorhinocladiella-like conidiophores.

Some of the synnemata-forming Ophiostoma species with ITS sequences differing greatly from those of O. piceae include O. ips, O. simplex, O. galeiformis and O. cucullatum. These species lack a Sporothrix anamorph, though O. ips forms micronematous, Hyalorhinocladiella-like conidiophores (Hoog 1993). Many of these and other synnemata-forming Ophiostoma species without Sporothrix synanamorphs form ascospores with sheath-like outer walls, extending to a cucullate brim around the spores (e.g., O. clavigerum, O. simplex, O. galeiformis and O. cucullatum) or in a box shape (O. ips and O. ainoae). In contrast, reniform ascospores without thickened outer cell walls are typical for species in the O. piceae complex.

The genus Pesotum was defined by the presence of both synnemata and micronematous conidiophores with denticles, with the synanamorphs of Ophiostoma ulmi as the type species (Crane and Schoknecht 1973). Accepting their definition, we would restrict Pesotum to those anamorphs with affinities to the O. piceae complex. Other synnemata anamorphs in Ophiostoma may have arisen through convergent evolution, perhaps from Leptographium-type conidiophores. The genus Phialographium is available for synnematos anamorphs of Ophiostoma species (e.g., O. sagmatosporae and O. cucullatum) with phialidic conidiogenous cells.

Species delimitation and identification in the O. piceae complex.—Use of a wide array of isolates from around the world and a holistic approach, including mating reactions, phylogenetic analysis, and phenotype (morphological and physiological), allows for a clearer delimitation of the species in the O. piceae complex. We emphasize phenotype in delimiting species (Harrington and Rizzo 1999), but mating compatibilities and phylogenetic analyses prove valuable in identifying lineages or mating populations where unique phenotypic characters may be found. It is noteworthy that ITS sequences were unable to distinguish O. piceae from O. canum, two species that are morphologically and biologically distinct. Other studies (Harrington and Rizzo 1999, Witthuhn et al 2000) have also found that ITS sequences of sympatric sibling species may be identical, and a strictly genealogical species concept cannot be applied if only nuclear rDNA sequences are utilized.

Production of the sexual state in culture is relatively easy for species in the O. piceae complex, and the heterothallic nature of most of the species allows for the identification of mating populations or biological species. Partial interfertility between species, as evidenced by poor ascospore production or aborted asci (Brasier 1993, Harrington and McNew 1998) was seen in some pairings between different species, and this strongly suggests that the species are distinct. In pairings between isolates of the same species, we generally had more success in producing perithecia and ascospores by spermatising a recipient (female) strain with conidial suspensions of a donor (male) strain than by simply pairing two strains together on a agar medium. Two mating types are seen in O. ulmi, O. novo-ulmi, O. himal-ulmi (Brasier and Mehrrota 1995), O. piceae, O. querici, (Brasier 1993), O. setosum (Uzunovic et al 2000), and O. flocosum, but Goidanich (1935) reported O. catoniamum to be homothallic.

A number of physiological and morphological characters were found useful in identifying cultures to species (TABLE II). Many of these characters have been used by others (Brasier and Kirk 1989, 1993, Brasier and Stephens 1991, 1993, Hamschlager et al 1994, Pipe et al 1995, Webber and Brasier 1990) to distinguish O. piceae and O. querici. Some overlapping morphological characters have been used to distinguish these two species, and these characters, growth at 32 C, and mating reactions with testers are sufficient to identify species in the complex, but a more rapid and objective identification criterion is the RFLP patterns of the amplified ITS region.

We did not attempt to distinguish O. ulmi, O. novo-ulmi, and O. himal-ulmi by the PCR-RFLP technique, although ITS sequence divergence should allow de-
velopment of a similar protocol for distinguishing these three species. More importantly, the species most commonly found on conifer sapwood (O. piceae, O. floccosum, O. querci and O. setosum) can be identified by ITS-RFLP patterns. With experience, it is possible to amplify the ITS region using scrapes of mycelium for template DNA. Kim et al (1999) showed that the ITS region of species in the O. piceae complex can also be amplified directly from conidia at the apex of synnemata, and primers were designed to distinguish O. piceae and O. querci. With the RFLP diagnostic technique employed here, more species can be distinguished. Without the need for DNA extraction, the PCR-RFLP technique can be completed in a single day, as has been demonstrated for members of Armillaria (Harrington and Wingfield 1995).

Phylogenetic analysis of the ITS region supports the distinction between O. piceae and O. querci, with O. piceae grouping with other species (O. canum, O. floccosum, and O. setosum) found primarily on conifers. In contrast, O. querci has been reported primarily from hardwoods in Europe, as have its relatives, O. catonianum and the Dutch elm disease fungi. However, O. querci is apparently common on sapwood of Pinus and other Pinaceae in North America and the Southern Hemisphere.

Conifer-inhabiting species of the O. piceae complex.—Ophiostoma canum, O. floccosum, and O. setosum have only been isolated from conifers and are more closely related to O. piceae than to the species found primarily on hardwoods. The ITS sequence of O. canum is not distinguishable from that of O. piceae, but lack of interfertility with O. piceae mating testers and the distinctive conidia of O. canum set this species apart. Partial interfertility with O. piceae suggests that both of our isolates of O. canum are mating type B; thus, we do not appear to have the opposite mating type for production of perithecia and ascospores in culture. The other conifer-inhabiting species appear to be common sapwood staining fungi not associated with any particular insect vector, but O. canum may be an important symbiont with a specific insect (the bark beetle Tomicus minor, Mathiesen 1950). The collection of thick-walled conidia at the apex of the synnema may be an adaptation for mycophagy by the bark beetle.

Our primary interest in the O. piceae complex has been in those species that cause bluestain in conifer wood. The most common synnema-forming species we have encountered in New Zealand and western North America are O. piceae, O. floccosum, O. setosum, and O. querci. It is relatively common to see two or more of these species on a single chip of wood, and mixed cultures of these species are very frequently seen, so these species apparently grow together well, with little inhibition of each other. Such mixed cultures have likely contributed to the confusion over the identification of these species. Unfortunately, the work done on O. piceae as a bluestain fungus (as reviewed by Seifert 1993) is difficult to interpret now that we realize that the studies could have utilized O. querci, O. floccosum or O. setosum.

Hardwood-inhabiting species of the O. piceae complex.—The taxonomy of O. querci remains unclear, and the basionym for the species is not ideal. No Latin description, dried specimen or culture was provided by Georgévitch (1926, 1927), but the fact that he described the fungus from oak in Europe (Croatia) makes it likely that his concept of the species matches the modern concept of the “hardwood form” of O. piceae. Accepting this, Loos’ (1932) species, O. fagi, is a clear synonym. We do not have authentic material for O. roboris, O. kubanicum, and O. valachicum, but these, too, are likely synonyms of O. querci. If O. roboris is a synonym, then Graphium roboris would be the earliest available name for the synnematus anamorph of O. querci. Okada et al (1998) proposed Pesiota pirinum as the anamorph of O. querci, but this is the anamorph name for Ophiostoma catonianum, which we consider distinct from O. querci. We also examined isolates from New Zealand and Vietnam that looked similar to O. querci but differed slightly in ITS sequence, and these cultures did not mate with O. querci testers. We suspect that more species closely related to O. querci will be recognized in the future.

The relatedness of O. querci to the Dutch elm disease fungi O. ulmi and O. novo-ulmi was demonstrated by similarity of ITS sequences. These Dutch elm disease fungi are very closely related but distinct species that have been introduced to North America and Europe. They may be native to Asia, though they have not been found there (Brasier 1990, Brasier and Mehrotra 1995). Ophiostoma himal-ulmi does appear to be native to the Himalayas, where it was found associated with bark beetle attacks in elm branches (Brasier and Mehrotra 1995). In inoculation studies, O. himal-ulmi was able to cause a vascular wilt disease in elm seedlings, and it was thought that this species was closely related to O. ulmi and O. novo-ulmi (Brasier and Mehrotra 1995). However, the ITS sequence analysis suggests that O. himal-ulmi is more closely related to O. querci.

The capacity of O. querci to cause disease in hardwood species has been debated, and fungi similar to O. querci have been frequently associated with declining oak trees in Europe (Brasier 1990, Brasier and Kirk 1989, Brasier and Mehrotra 1995, Harrington
1993, Morelet 1992, Oleksyn and Przybyl 1987). Its close relationship to *O. himal-ulmi*, *O. ulmi*, and *O. novo-ulmi* also suggests that it has the potential to cause a vascular disease, and attention should be given to introduction of this species to new environments.

We suspect that most species in the *O. piceae* complex are native to the Northern Hemisphere. Indeed, the common occurrence of *O. querci* on sapwood of Pinaceae in some regions (such as the Southern Hemisphere and North America) may be due to its not being native to these ecosystems. This species and others in the complex will produce synnemata and/or perithecia on logs, crating, dunnage and other materials commonly discarded from ships, and we believe that the wide distribution of some of these species is due to human activities.

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